# Protein-protein surface interactions: Constrains of homologous versus heterogeneous domains

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#### ABSTRACT

Interactions of proteins could be resulted from homodimerisation or heterodimerisation. But these are quite specific and always selective in nature and act only on particular set of class of proteins, which provide substantial target behaviour to a drug designer. Here in this paper, provided a commentary based review compiling the short notes information provided on protein proteins interactions, by "Catherine Royer"

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### INTRODUCTION

Interactions of biomolecules lead to the various biochemical events. Even drugs derived from natural source, as well as synthetics, also needs to interact with biomolecules to affect their activity<sup>[1-3]</sup>. Although protein-protein interactions (PPIs) are the keys, to regulate or transduce a signal to a function for maintaining a normal homeostasis, but any fault leads to a malfunctioning and diseases (say e.g. anaphylactic shock)<sup>[4, 5]</sup>. Around > 99% metabolic routes utilised the PPIs for tight hold. In recent years, various genomic tools and techniques <sup>[6, 7]</sup> were used to understand the nature of PPIs and now it is much established that these interactions can often result from a physical contacts, involving bonding and electrostatic forces.

PPIs are critical for intracellular signalling, endocytosis and internalisations. They are crucial to the understanding of all in vivo functions, cellular regulation, biosynthetic and degradation pathways, signal transduction, Nucleic acid replication, transcription and translation, multi-molecular assembly associations. cellular packaging, optimisation of immune response. They also relate to allosteric mechanisms, either turning genes on or off and therefore attain high interest for drug designing. All biological processes are regulated through association and dissociation of protein molecules. These includes endogenous biomolecules receptor binding, protease inhibition of protein lysis, Antibody-antigen recognition efficacy, signal transduction mechanisms, enzyme substrate affinity, vesicle transport, Post-translations or -transcriptions modifications. Hence, PPIs are now highly targeted and become centre stage of protein science. The ability to predict the preferred method by which proteins interact would facilitate assignment of protein function.

#### **CLASSES OF PROTEIN-PROTEIN INTERACTIONS**

Majorly PPIs resulted into the complexes formation, as homo-oligomers or hetero-oligomers. The homooligomer are macromolecular complexes constituted by only one type of protein subunit. Protein subunit assembly is guided by the establishment of noncovalent interactions in the quaternary structure of the protein. Disruption of homo-oligomers in order to return to the initial individual monomers often requires denaturation of the Complex. Several enzymes, carrier proteins, scaffold proteins, and transcriptional regulatory factors carry out their functions as homo-oligomers. Distinct protein subunit interacts in hetero-oligomers, which are essential to control several cellular functions. The importance of the communication between heterologous protein is even more evident during cell signaling events and such interactions are only possible due to structural domains within the proteins.

There is another category where they are considered their interactions in stability context. **Stable interactions** involve proteins that interact for a long time, taking part of permanent complexes as subunits, in order to carry out structural or functional roles. These are usually the case of homooligomers (e.g. cytochrome c), and some heterooligomeric proteins, as the subunit of ATPase. On the other hand, a protein may interact briefly and in a reversible manner with other proteins in only certain cellular contexts-cell type, cell cycle stage, external factors, presence of other binding proteins etc. These are called transient interactions (e.g.- Could be seen in 5HT receptor-macromolecular complexes <sup>[8]</sup>). The way to classify these interactions as covalent and noncovalent interactions. Covalent interactions are those with the strongest association and are formed by disulphide bonds or electron-sharing. Although being rare, these interactions are determinant in some posttranslational modifications, as ubiquitination and sumoylation. Noncovalent interactions are usually established during transient interactions by the combination of weaker bonds, such as hydrogen bonds, ionic interactions, van der Waals forces, or hydrophobic bonds.

#### **Homo-oligomerization**

The noncovalent interactions of protein quaternary structure provide, an edge over the cases where the functional unit comprised of a linear polypeptide chain. Initial binding of ligand excels energy on subunit site of protein and drives it to adopt an alter transition conformation. However, the modulation of subunit affinity towards the ligand binding is independent of the adopted fold structure of the protein, providing a considerable energetic margin for modulation of activity. Also, these experiments told the protein regulatory functions works up to an appropriate molar concentration range and later the affinity of protein subunit fades out, indicating the role of protein concentration dependent. These



**Figure 1**. Homo oligomerization of haemoglobin tetramer (PDB: 1GZX)<sup>[9]</sup>

regulatory functions can either activate or inactivate the progression of cellular any process induced by oligomer or the monomer. Hence it is worth to say that the function in

oligomers

can fine-tuned by available ligand concentration (could any ions, ortho- or allosteric substrate and ligands etc.) and by protein concentration (comparative level of expression or degradation rates), see figure as an example of homooligomerisation.

#### Interactions between heterologous proteins

More often the heterologous protein interactions came to known and become more popular when it came apparent that physical state or the ambient environmental affects the cellular metabolic rate (like these interacting proteins can be affected by the chemical presence, change in luminosity etc). However later math and current research found them more valuable and critical in various other, metabolic and much required homeostasis functions. Although here we encompass below some of the heterologous proteins.

#### Protein domains involved in Signal Transduction

*Src* homology domains 2 and 3, are commonly referred to as *SH2* and *SH3* domains. The SH2 domains recognize tyrosine phosphorylated proteins (particular autophosphorylated growth factor receptors) <sup>[10]</sup>. A ribbon cartoon shown below, indicating the *SH2* domain of the lymphocyte specific tyrosine kinase, P56<sup>*lck*</sup> along a co-crystallise peptide <sup>[11]</sup>, see figure 2. The structural architecture of *SH2* domains possess of a three-stranded twisted beta sheet which is sandwiched between two alphahelices. The *SH2* domain also have a deep cleft which acquired high affinity for phosphotyrosine, but not for, either phosphoserine or phosphothreonine.



Figure 2. Ribbon diagram of the SH2 domain from  $p56^{lck}$  (green ribbons) bound to phosphopeptide (spacefilled cyan) (PDB: 1CWE).

Residues starting for C-terminal to up phosphotyrosine, has been noticed as an active site to confer the specificity of the interaction. Whereas the second pocket assist in recognizing these Cterminal specificity determinants. Various classes of SH2 domains recognize numerous types of Cterminal sequences (Class I SH2 domains recognize Cterminal tripeptides P-P-H, where P and H stand for polar and hydrophobic; Class III SH2 domains recognize H-X-H tripeptides). Till this date, number of SH2 domains three dimensional structures have been resolved and found a clear evident of structural homology (predicted by the comparative analysis of SH2 domain from p561ck and C-terminal SH2 domain of the Syk tyrosine kinase) [12].

Like SH2 domains, SH3 domains are also found in similar signal transducer proteins (protein tyrosine kinases), however they recognize polyproline type II helical structures (PXXP motifs) in their binding proteins (e.g., SH3 domain of Grb2 recognizes the PXXP motif of a guanyl nucleotide releasing factor; SH3 domain of the src kinase, Fyn, can bind the PXXP motif of the HIV-1 protein Nef; SH3 domain of fyn proto-oncogene complexed with a synthetic p85 subunit peptide of Phosphoinositide 3-kinase) <sup>[13]</sup> is shown in Figure 3. It could be easily seen that the SH3 domain architecture possesses of a beta barrel formed by two orthogonal beta sheets, which is constructed of three anti-parallel beta strands and the proline rich site on the partner proteins pack onto aromatic residues of the SH3 domains.



**Figure 3.** NMR Structure of the *SH3* Domain from Fyn Proto-Oncogene Tyrosine Kinase complexed with the synthetic peptide P2I corresponding to residues 91-104 of the P85 Subunit of PI3-Kinase, (PDB 1AZG)

PDZ domains is an another class of protein interaction motifs which has GLFG repeats or DHR domains and were identified initially in three proteins, PSD-95, DlgA and ZO-1 (therefore they called so, "PDZ domains") where all are guanylate kinases. These are highly involved in ion channel receptor clustering, receptor/enzyme coupling and a variety of other protein assembly associations. The PZD domains recognize *C*-terminal tri-peptide motifs (S/TXV), other PZD domains or even LIM domains (see in Figure 4). Surprising resemblance of PDZ domain of DgIA with SH2 domain, tells similar structural architecture including a beta barrel sandwiched between two alpha-helices. The below figure shows the structural PDZ domain of PSD95 holding tightly the *C*-terminal peptide from Cript <sup>[14]</sup>.



**Figure 8.** PDZ domain of PSD-95 complexed with the C-terminal peptide derived (PDB 1BE9)

The another cellular signalling domain is the LIM domain. These domains are cysteine enriched zinc finger domains present in many homeodomain and non-homeodomain proteins involved critically in cellular development, differentiation, association with the cyto-skeletons or in cellular senescence. These proteins need PDZ motifs, bHLH transcription factors (see below), other LIM domains, and LIM binding proteins to bind. These were initially found in 3 homeodomain transcription factors (lin11, isl1 and mec3, hence LIM domain), which holds the consensus sequence CX2CX16-23HX2CX2CX2CX16-21CX2C/H/D, and now identified in over 35 proteins to date. The NMR structure of the LIM domain from the LASP protein was first identified in 1996 <sup>[15]</sup>. The zinc atom (in space-filled cyan colour), coordinated by cysteine residues in the two loops corresponding to the double finger, also shown in figure 5.



**Figure 5**. Structure of the amino terminal of LIM domain peptide of the LASP protein (PDB: 1ZFO)

The **pleckstrin homology domain** (also called, "PTB domains") proteins also transduce signal in cellular intracellular signalling. These proteins bind to acidic domains in signal transducer proteins as well as to phosphoinositdes, the WW domain, a semiconserved region of 38-40 amino acids, containing two conserved tryptophan at 20 amino acid spaced and hence called so as "WW" (WSxWS motif in cytokine receptors and the WD repeat).

# PROTEIN DOMAINS INVOLVED IN TRANSCRIPTIONAL REGULATION

The nucleic acid protein juncture has been highlighted since when it gets focused by the researcher for the first time, especially modulation of the gene expression gets favouritism from the researchers in 21<sup>st</sup> century. Although these processes require selective recruitment of specific proteins, selective energy based conformation for the affinity and hence highly structural specific. Numerous prokaryotic and eukaryotic transcription factors were found by biochemists with or without their key role in the cell. For better understanding, 3D structures were characterised and classified further, based on sequence or structural homology, except some instances where they found on functional homology. One of highly focused family is that of the **bHLH** transcription factors, so called as its structural motif include a basic region for binding DNA and a helixloop-helix region involved in dimerization. This family include the MyoD family of transcription factors which regulate the gene expression in muscle cells and control their differentiation to mature muscle cells. Other members of the bHLH family are the ubiquitous products of the E2A gene, E12 and E47.

Although MyoD family members can form homo as well as heterodimers with the E12 and E47 proteins, see figure 6.



Figure 6. bHLH domain of MyoD complexed with DNA (PDB 1MDY)

Another class includes leucine zipper transcription factors, c-jun and c-fos (constituents of the AP1 transcription factor). AP1 is highly involved in gene transcription regulation in cell proliferation. Similar to bHLH family, bzip family members also interacts by their coiled alpha helices. Interestingly, the bzip proteins possess the leucine residues positioned at helix turn providing ample hydrophobic interface for the PPIs, hence so called as leucine zipper. Nterminal to the zipper recognize and bind to specific DNA sequences. These family members also homoand hetero-dimerize together or other family protein and can alter the DNA specificity and the transcriptional rate (see in figure: b-zip domains of the c-Fos/cJun heterodimer bound to DNA, see in Figure 7) <sup>[16]</sup>.



**Figure 7.** Schematic representation of c-Fos/c-Jun heterodimer complexed with DNA (PDB 1FOS)

**The RHR or Rel Homology** Region containing transcription factors are another class that are functional active in cellular immune responses or further de-differentiation. NF- $\kappa$ B, is one of the key member of this class and has specificity to DNA target site in the immunoglobulin light chain enhancer and also regulating various gene expression in response to invading infection. The **Rel Homology** Region of the two subunits of NF- $\kappa$ B (p50 and p65 or RelA) specifies DNA binding, dimerization and nuclear localization. Crystallographic data revealed RHR region in the p50 subunit of NF- $\kappa$ B <sup>[17]</sup> resembled like beta sandwich structure of the immunoglobulin folds of immunoglobulins and also assist in the dimerization with DNA.

**Ankyrin repeat** are ubiquitous domains and found in more than 100 different proteins of diverse functionality like catalytic, transcription or inhibitors of transcription activities similar-like *lk*B, that inhibits the NF-kB transcription factor. Ankyrin repeats contain around 32-33 amino acids and usually proteins hold at least 4 copies of this domain. The structural frame processes beta strand, helix turn helix extended strand beta strand segment. Whereas the helices and beta strand are anti-parallel to each other and the plane of the strand regions is perpendicular to the helical axis (See Figure 13 below). The consensus sequence of the ankyrin repeat is:

- G – T P L H L A A R – G H V E V V K L L L D – G A D V N A – T KA I S Q N N LD IA E V K N P D DV K T M R Q S I NE

Another large family includes the family of steroid/nuclear receptors including steroidal hormone receptors, vitamin D receptor, retinoic acid receptor (RAR and RXR) and thyroid hormone receptors, and a several other orphan receptors and transcription factors. The structural construction is quite unique of these receptors comprising Nterminal constitutive and tissue specific transcriptional activation domain (TAF1) which can have varied in length, central region for DNA binding and dimerization domain (DBD, linker region for nuclear localization and C-terminal as ligand binding domain (LBD) that also holds ligand dependent transcriptional activation activity (TAF2) and dimerization determinants. However, the structural information of the DBD and LBD of these receptors drawn interest of putative drug targets, but also challenged the drug designer as they share high degree of structural homology in their structures. The individual DBD homodimerizes with DNA through palindromic response elements (HRE's) and shows cooperativity relationship for the binding of the other DBD of the protein on DNA.



Figure 8. Structure of the ERDBD bound to DNA

It was also seen that these DNA Binding Domains (DBD) have highly conserved pair of Zn fingers, one of which involves in DNA recognition helix, and the other holds the determinants for dimerization. However, the D-loop participate in the dimer interface, flanked by two cysteine residues which coordinates with Zn atom (see in

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## **↓** REFERENCES

1. Negi A, Gill B: Success stories of enolate form of drugs. PharmaTutor 2013, 1(2):45-53.

2. Singla R, Negi A, Singh V: Indole based alkaloid in cancer: an overview. PharmaTutor Mag 2014, 2:76-82.

3. Singla R, Singh V, Negi A: Synthetic Indole Alkaloids in Cancer: An Overview.

4. Negi A: Anaphylactic Shock: Shocking Error of Immune System! PharmaTutor 2013, 1(2):12-16.

5. Negi A, Pandey AK, Joshi G, Agnihotri V: Impact of protein tyrosine phosphate on cancer metastasis: an overview. World J Pharm Res Technol 2013, 1(2):118-130.

6. Negi A, Gill B, Anand S: Tilling: versatile reverse genetic tool. PharmaTutor 2014, 2(1):26-32.

7. Franceschini A, Szklarczyk D, Frankild S, Kuhn M, Simonovic M, Roth A, Lin J, Minguez P, Bork P, von Mering C: STRING v9. 1: protein-protein interaction networks, with increased coverage and integration. Nucleic acids research 2013, 41(D1):D808-D815.

8. Zhou J, Reidy M, O'Reilly C, Jarikote DV, Negi A, Samali A, Szegezdi E, Murphy PV: Decorated Macrocycles via Ring-Closing Double-Reductive Amination. Identification of an Apoptosis Inducer of Leukemic Cells That at Least Partially Antagonizes a 5-HT2 Receptor. Organic letters 2015, 17(7):1672-1675.

9. Paoli M, Liddington R, Tame J, Wilkinson A, Dodson G: Crystal structure of T state haemoglobin with oxygen bound at all four haems. Journal of molecular biology 1996, 256(4):775-792.

10. Negi A, Ramarao P, Kumar R: Recent Advancements in Small Molecule Inhibitors of Insulin–like Growth Factor-1 Receptor (IGF-1R) Tyrosine Kinase as Anticancer agents. Mini reviews in medicinal chemistry 2013, 13(5):653-681.

11. Tong L, Warren TC, Lukas S, Schembri-King J, Betageri R, Proudfoot JR, Jakes S: Carboxymethyl-phenylalanine as a replacement for phosphotyrosine in SH2 domain binding. Journal of Biological Chemistry 1998, 273(32):20238-20242.

12. Narula S, Yuan R, Adams S, Green O, Green J, Philips T, Zydowsky L, Botfield M, Hatada M, Laird E: Solution structure of the C-terminal SH2 domain of the human tyrosine kinase Syk complexed with a phosphotyrosine pentapeptide. Structure 1995, 3(10):1061-1073.

13. Renzoni D, Pugh D, Siligardi G, Das P, Morton C, Rossi C, Waterfield M, Campbell I, Ladbury J: Structural and thermodynamic characterization of the interaction of the SH3 domain from Fyn with the proline-rich binding site on the p85 subunit of PI3-kinase. Biochemistry 1996, 35(49):15646-15653.

14. Doyle DA, Lee A, Lewis J, Kim E, Sheng M, MacKinnon R: Crystal structures of a complexed and peptide-free membrane protein–binding domain: molecular basis of peptide recognition by PDZ. Cell 1996, 85(7):1067-1076.

15. Hammarström A, Berndt KD, Sillard R, Adermann K, Otting G: Solution structure of a naturally-occurring zinc-peptide complex demonstrates that the N-terminal zinc-binding module of the Lasp-1 LIM domain is an independent folding unit. Biochemistry 1996, 35(39):12723-12732.

16. Glover JM, Harrison SC: Crystal structure of the heterodimeric bZIP transcription factor c-Fos–c-Jun bound to DNA. 1995.

17. Ghosh G, Van Duyne G, Ghosh S, Sigler PB: Structure of NF-κB p50 homodimer bound to a κB site. 1995.

18. Schwabe JW, Chapman L, Finch JT, Rhodes D: The crystal structure of the estrogen receptor DNA-binding domain bound to DNA: how receptors discriminate between their response elements. Cell 1993, 75(3):567-578.